



AssayMax Human Fibronectin (FN) ELISA Kit

Catalog Number EF2045-1

Introduction

Fibronectin (FN) is a major component of the extracellular matrix and blood plasma, and is a specific ligand for several integrin adhesion receptors (1). FN plays an important role not only in cell adhesion (2) and wound healing (3), but also in embryogenesis (4) and hematopoiesis (5). FN is over-expressed in cardiovascular disease states such as atherosclerosis (6) and myocardial infarction (7). Reduced levels of FN have been reported in patients with Disseminated Intravascular Coagulation (DIC) and low concentrations appear to correlate with a poor prognosis (8).

Principal of the Assay

The AssayMax Human Fibronectin ELISA kit is designed for detection of human FN in cell culture media and urine samples. This assay employs a quantitative sandwich enzyme immunoassay technique that measures human FN in 3.5 hours. A polyclonal antibody specific for human FN has been pre-coated onto a 96-well microplate with removable strips. FN in standards and samples is sandwiched by the immobilized polyclonal antibody and biotinylated polyclonal antibody specific for human FN, which is recognized by a streptavidin-peroxidase conjugate. All unbound material is then washed away and a peroxidase enzyme substrate is added. The color development is stopped and the intensity of the color is measured.

Caution and Warning

- This kit is for research use only.
- The kit should not be used beyond the expiration date.
- The Stop Solution is an acid solution.
- All human source materials have been tested and found to be negative to HbsAg, HIV-1 and HCV by FDA approved methods.

Reagents

- **FN Microplate:** A 96-well polystyrene microplate (12 strips of 8 wells) coated with a polyclonal antibody against FN
- **Sealing Tapes:** Each kit contains 3 pre-cut, pressure-sensitive sealing tapes that can be cut to fit the format of the individual assay.

- **FN Standard:** Human FN in a buffered protein base (1 µg, lyophilized)
- **Biotinylated Human FN Antibody (100x):** A 100-fold concentrated biotinylated polyclonal antibody against human FN (80 µl).
- **Streptavidin-Peroxidase Conjugate (SP Conjugate):** A 100-fold concentrate (90 µl)
- **MIX Diluent Concentrate (10x):** A 10-fold concentrated buffered protein base (30 ml)
- **Wash Buffer Concentrate (10x):** A 10-fold concentrated buffered surfactant (2 x 30 ml)
- **Chromogen Substrate:** A ready-to-use stabilized peroxidase chromogen substrate tetramethylbenzidine (8 ml)
- **Stop Solution:** A 0.5 N hydrochloric acid (12 ml) to stop the chromogen substrate reaction

Storage Condition

- Store unopened kit at 2-8⁰C up to expiration date.
- Opened reagents may be stored for up to 1 month at 2-8⁰C. Store reconstituted standard and Biotinylated FN at -20⁰C or below.
- Opened unused strip wells may return to the foil pouch with the desiccant pack, reseal along zip-seal. May be stored for up to 1 month in a vacuum desiccator.

Other Supplies Required

- Microplate reader capable of measuring absorbance at 450 nm
- Pipettes (1-20 µl, 20-200 µl, and multiple channel pipettes)
- Deionized or distilled reagent grade water

Sample Collection, Preparation and Storage

- **Cell Culture Supernatants:** Centrifuge cell culture media at 2000 x g for 10 minutes to remove debris. Collect supernatants and assay. Store samples at -20⁰C or below. Avoid repeated freeze-thaw cycles.
- **Urine:** Collect urine using sample pot. Centrifuge samples at 800 x g for 10 minutes and assay. Dilute samples 1:2 into MIX Diluent. Store undiluted samples at -20⁰C or below for up to 3 months. Avoid repeated freeze-thaw cycles.

Reagent Preparation

- Freshly dilute all reagents and bring all reagents to room temperature before use. If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved.
- **Standard Curve:** Reconstitute the 1 µg of FN Standard with 1 ml of MIX Diluent to generate a 1 µg/ml of solution. Allow the standard to sit for 10 minutes with gentle agitation prior to making dilutions. Prepare triplicate standard points by serially diluting the standard solution (1 µg/ml) 1:4 with MIX Diluent to produce 0.25, 0.063, 0.016 and 0.004 µg/ml solutions. MIX Diluent serves as the zero standard (0 µg/ml). Any remaining solution should be frozen at < -20⁰C.

Standard Point	Dilution	[FN] ($\mu\text{g/ml}$)
P1	Standard (1 $\mu\text{g/ml}$)	1.000
P2	1 part P1 + 3 parts MIX Diluent	0.250
P3	1 part P2 + 3 parts MIX Diluent	0.063
P4	1 part P3 + 3 parts MIX Diluent	0.016
P5	1 part P4 + 3 parts MIX Diluent	0.004
P6	MIX Diluent	0.000

- **Biotinylated Human FN Antibody (100x):** Spin down the antibody briefly and dilute the desired amount of the antibody 1:100 with MIX Diluent.
- **MIX Diluent Concentrate (10x):** Dilute the MIX Diluent 1:10 with reagent grade water.
- **Wash Buffer Concentrate (10x):** Dilute the Wash Buffer Concentrate 1:10 with reagent grade water.
- **SP Conjugate (100x):** Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 1:100 with MIX Diluent.

Assay Procedure

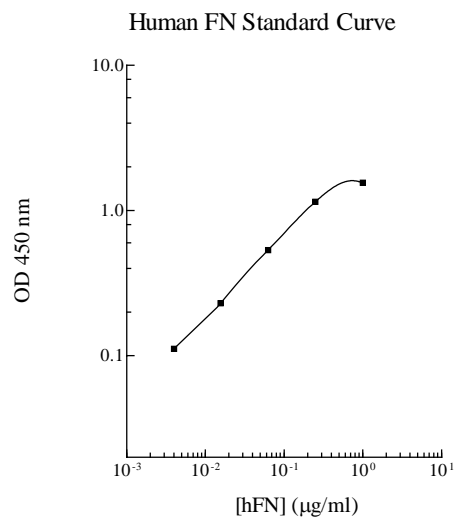
- Prepare all reagents, working standards and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at room temperature (20-30°C).
- Remove excess microplate strips from the plate frame and return them immediately to the foil pouch with desiccant inside. Reseal the pouch securely to minimize exposure to water vapor and store in a vacuum desiccator.
- Add 50 μl of standard or sample per well, and cover wells and incubate for 2 hours. Start the timer after the last sample addition.
- Wash five times with 200 μl of Wash Buffer. Invert the plate and decant the contents, and hit it 4-5 times on absorbent paper towel to completely remove liquid at each step.
- Add 50 μl of Biotinylated Human FN Antibody to each well and incubate for 60 minutes.
- Wash five times with 200 μl of Wash Buffer.
- Add 50 μl of Streptavidin-Peroxidase Conjugate to each well and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.
- Wash five times with 200 μl of Wash Buffer.
- Add 50 μl of Chromogen Substrate per well and incubate for about 10 minutes or till the optimal blue color density develops. Gently tap plate to ensure thorough mixing and break the bubbles in the well with pipette tip.
- Add 50 μl of Stop Solution to each well. The color will change from blue to yellow.
- Read the absorbance on a microplate reader at a wavelength of 450 nm **immediately**. Please note that after the reaction is stopped for about 10 minutes, some black particles may be generated at high concentration point, which will reduce the readings.

Data Analysis

- Calculate the mean value of the triplicate readings for each standard and sample.
- To generate a Standard Curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance on the y-axis. The best-fit line can be determined by regression analysis using 4-parameter or semi-log curve fit.
- Determine the unknown sample concentration from the Standard Curve and multiply the sample value by the dilution factor.

Standard Curve

- The curve is provided for illustration only. A standard curve should be generated each time the assay is performed.



Precision, Sensitivity and Specificity

- The minimum detectable dose of FN is typically 4 ng/ml.
- Intra-assay and inter-assay coefficients of variation were 5.0% and 7.0% respectively.

Recovery

Standard Added Value	0.02 – 0.2 µg
Recovery %	90-112 %
Average Recovery %	101 %

Linearity

Sample Dilution	Average Percentage of Expected Value
	Urine
1:2	101 %
1:4	98%
1:8	102%

Note: This kit can be used for rat or mouse plasma/serum samples. The suggested dilution is 1:100. The FN value in rat/mouse plasma/serum = 15 * calculated value from standard curve.

References

1. Hynes, R.O. (1992) *Cell* 69:11
2. Wu, C. *et al.* (1995) *Cell* 83:715
3. Brown, L.F. *et al.* (1993) *Am. J. Pathol.* 142:793
4. Pagani, F. *et al.* (1991) *J. Cell Biol.* 113:1223
5. Verfaillie, C.M. *et al.* (1991) *J. Exp. Med.* 174:693
6. Glukhova M.A. *et al.* (1989) *J. Cell. Biol.* 109:357
7. Knowlton, A.A. *et al.* (1992) *J. Clin. Invest.* 89:1060
8. Cembrowski, G.S. and Mosherb, D.F. (1984) *Thrombosis Research* 36:437

Revision 2.1